



Strong impact of temperature on hatching success of aquatic invertebrates dispersed by two waterbirds

Maria Bisquert-Ribes · Bárbara Alvado · Mireia Costa-Bisquerra · Alba Murria-Fernández · Pablo Vera · Francesc Mesquita-Joanes · Xavier Armengol

Received: 14 January 2025 / Revised: 20 June 2025 / Accepted: 23 June 2025
© The Author(s) 2025

Abstract Waterbirds are key agents of passive dispersal for aquatic invertebrates, particularly through internal transport of their propagules. In this study, we assessed the potential dispersal of invertebrates through endozoochory by two Anatidae species, the mallard and the common shelduck, in Albufera de Valencia Natural Park. We aimed at comparing the hatching success of invertebrate propagules egested by both ducks, under two temperatures, simulating cold (15 °C) and warm (25 °C) conditions. We performed a controlled hatching experiment using faecal

samples (30 per species) collected from the roosting areas of both ducks. A total of 34 invertebrate taxa hatched, including rotifers, cladocerans, copepods, ostracods, ciliates, and microturbellarians. Species richness and hatching rates were higher under warm conditions, and shelduck samples showed more diverse assemblages than mallards. Five of the detected taxa are considered invasive non-native to the region. Some of the rotifer and ostracod species found, as well as microturbellarians, represent first records of internal dispersal by waterbirds. Our findings suggest that temperature can influence hatching success and may shape the community composition of colonizing organisms through endozoochory. Moreover, Anatidae can be effective vectors for aquatic invertebrates, including non-native species, with potential implications for metacommunity dynamics and invasions in Mediterranean wetlands.

Guest editors: Janet Higuti, Claudia C. Bonecker, André A. Padial, Stuart Halse & Jonathan Rosa / Ecology and Evolution of Dormancy and Diapause in Aquatic Organisms.

Supplementary Information The online version contains supplementary material available at <https://doi.org/10.1007/s10750-025-05936-9>.

M. Bisquert-Ribes (✉) · M. Costa-Bisquerra · A. Murria-Fernández · F. Mesquita-Joanes · X. Armengol
Cavanilles Institute of Biodiversity and Evolutionary Biology, University of Valencia, Carrer Catedràtic José Beltrán Martínez, 2, 46980 Paterna, Spain
e-mail: maria.bisquert@uv.es

M. Costa-Bisquerra
e-mail: mireiacosta97@gmail.com

A. Murria-Fernández
e-mail: Almufer9510@gmail.com

F. Mesquita-Joanes
e-mail: francesc.mezquita@uv.es

X. Armengol
e-mail: javier.armengol@uv.es

B. Alvado
Image Processing Laboratory, University of Valencia, Carrer Catedràtic Agustín Escardino Benlloch, 9 E4 building – 4th Floor, 46980 Paterna, Spain
e-mail: Barbara.Alvado@uv.es

P. Vera
Servicio de Conservación de Ambientes Acuáticos, SAV-Servicio Devesa-Albufera, Valencia, Spain
e-mail: pablovera_g@hotmail.com

Keywords Endozoochory · Hatching experiment · Invasive species · Metacommunity · Resting propagules

Introduction

Metacommunities are structured by a variety of factors, including local environmental conditions, species interactions, spatial dynamics and temporal effects (Leibold & Chase, 2018). Dispersal, the main mechanism driving spatial dynamics, can control metacommunity structure by interchanging species and genetic material between sites (Gálvez et al., 2022), facilitating the movement of species between different habitats, which is essential for maintaining biodiversity. High dispersal rates can lead to homogenization of communities across sites (Heino et al., 2015), while limited dispersal can result in distinct community compositions based on local environmental conditions (Leibold et al., 2004; Heino et al., 2015).

Dispersal becomes especially relevant in aquatic ecosystems. While many aquatic organisms, including insects, amphibians, birds, reptiles, and mammals, can actively move between water bodies across land, others cannot do so, such as many invertebrates and plants. Instead, they rely on passive transportation via various vectors, whether abiotic, such as wind or waterflow (Hulsmans et al., 2007; Brendonck et al., 2010), or biotic, involving living organisms (Alekseev et al., 2007; Vanschoenwinkel et al., 2008, 2011; Sabagh et al., 2011). A wide variety of species can serve as vectors for aquatic invertebrates, ranging from fish, mammals and birds to humans themselves (Vanschoenwinkel et al., 2008, 2011; Waterkeyn et al., 2010; Valls et al., 2016).

Aquatic birds, in particular, are highly important for the dispersal of invertebrates due to their high mobility at both local and regional scales (Figuerola & Green, 2002; Green et al., 2002), playing a key role in ecosystem functioning (Green & Elmberg, 2014). While all aquatic birds can act as dispersers through epizoochory (or ectozoochory), waterfowl (Anatidae) have been identified as particularly relevant for endozoochory, facilitating the dispersal of plants (Kleyheeg et al., 2016; Soons et al., 2016; Lovas-Kiss et al., 2019) and invertebrates (Figuerola et al., 2003; Brochet et al., 2010; Valls et al., 2017; Moreno

et al., 2019). Moreover, zoochory by waterbirds has been documented for a variety of invasive non-native plants and aquatic invertebrates (Reynolds et al., 2015; Green, 2016) after their initial introduction by humans.

For a successful colonisation, propagules must be transported, and the species should be able to establish and grow to maturity in the habitats they arrive. Factors such as habitat suitability, propagule traits, and the effects of gut passage are critical in this process (Green et al., 2023). Waterbirds can facilitate directional dispersal into compatible environments and increase establishment chances for aquatic organisms with high propagule pressure, as well as those capable of clonal reproduction, such as certain cladocerans, ostracods, and rotifers, allowing colonization even from a small number of propagules (Green et al., 2016, 2023; van Leeuwen et al., 2023).

Environmental conditions should be suitable for the hatching of resting propagules to establish their populations. Several factors, such as salinity, photoperiod, pH or temperature can often trigger the hatching of these propagules (Alekseev et al., 2007). A previous study in a Mediterranean coastal wetland (Bisquert-Ribes et al., 2024) indicated that the microcrustacean (cladocerans, copepods and ostracods) metacommunity differs in species composition between the coldest season, winter, and the warmest season, summer. Both seasons showed the presence of typical native species, but invasive non-native (invasive, hereafter; Soto et al., 2024) ostracods were found with a higher prevalence in summer, suggesting temperature (and probably other environmental factors) may affect community variation, particularly in temporary habitats where the egg bank serves as the core of the community that re-establishes after the drying phase (Brendonck & De Meester, 2003; Olmo et al., 2020).

Here, we focused on the role of two waterfowl species as vectors for the dispersal of invertebrates in a coastal wetland of the Iberian Peninsula through a hatching experiment of resting propagules from faeces. Our main aim was to determine whether there was differential hatching of resting propagules under two different temperatures, simulating cold and warm conditions. We tested the following hypothesis: (1) the species composition and/or the hatching dynamics would vary between temperature treatments; (2) the two waterbird species would differ in the hatching

rates of the propagules they transport internally; (3) invasive invertebrate species, particularly ostracods and potentially cladocerans and copepods, would hatch, as they have been previously reported in the study area (Bisquert-Ribes et al., 2024); (4) invaders would appear more frequently under the warm temperature treatment, due to most of them being of tropical origin (McKenzie & Moroni, 1986; Bisquert-Ribes et al., 2023). These results may help clarify how temperature influences the emergence and the potential establishment of species whose propagules are dispersed in Mediterranean wetlands.

Material and methods

Study area and sampling methods

The Albufera Natural Park (listed as Ramsar and Natura 2000 site), spanning 21,120 ha, is a wetland complex situated on the Mediterranean coast of the Iberian Peninsula. This protected area consists of a shallow hypertrophic lake, surrounded by over 15,000 ha of rice fields and other aquatic systems such as marshes, freshwater springs, and temporary ponds. This area is remarkable within the broader context of European wetlands, particularly concerning aquatic birds, as it provides breeding, migratory stopover, and wintering habitats for a wide array of waterbirds. During the winter of 2021, over 66,000 waterbirds were recorded in the region. The most numerous groups included gulls (ca. 28,000), waterfowl (ca. 20,000), glossy ibises (ca. 11,600), and waders (ca. 5,000), among others (SEO BirdLife, 2021a).

The mallard (*Anas platyrhynchos* Linnaeus, 1758) is the most abundant Anatidae in the region, with over 8,000 individuals recorded in winter (SEO BirdLife, 2021a) and approximately 2,500 breeding pairs (SEO BirdLife, 2021b). The common shelduck (*Tadorna tadorna* Linnaeus, 1758), in contrast, is primarily a wintering species in the Albufera, exceeding 2,000 individuals in 2021 and with a small local breeding population restricted to protected areas like Racó de l'Olla (SEO BirdLife, 2021a; b). Seasonally, mallards remain in the region year-round but increase in number due to winter migrants arriving from northern and central Europe (Díaz et al., 1996; SEO BirdLife, 2012). Shelducks are mostly resident in southwestern Europe (Patterson, 1982), however, in the

Albufera, they arrive in autumn and reach their peak between December and February before leaving for other areas in March (SEO BirdLife, 2021a; b). Both species exhibit short-range dispersal within the area in response to habitat availability and disturbance. Shelducks show a preference for saline or brackish wetlands and mallards can be most frequently found in shallow, vegetated freshwater areas, in association with many different habitats of the Albufera (SEO BirdLife, 2021b).

In the context of our study, both species are relevant for their potential role in the passive endozoochorous dispersal of aquatic invertebrate propagules (e.g. Green et al., 2008; Valls et al., 2017; Lovas-Kiss et al., 2018). Their winter foraging behaviour, particularly in rice fields and marshes, involves techniques like dabbling, upending, head dipping, and sieving sediments (Liordos, 2010). These practices facilitate the ingestion of resting propagules directly from the sediment, plants and by the ingestion of adults carrying the propagules. Both species access prey items in shallow water and soft sediments, although shelducks often prefer saline and deeper environments for roosting (SEO BirdLife, 2021a).

Faecal samples were collected from resting/roosting areas of both duck species. Mallard samples were gathered at Tancat de l'Illa (39° 16' 41" N, 0° 17' 29" W). This is a managed freshwater wetland that functions as a green filter with nutrient-rich shallow waters and abundant submerged vegetation (Rodrigo et al., 2013a,b). In contrast, shelduck samples were obtained at Racó de l'Olla (39° 19' 54" N, 0° 19' 02" W), a restored brackish lagoon hydrologically isolated from the main lake and characterized by deeper waters and elevated salinity levels, habitat that is usually preferred by the shelduck (SEO BirdLife, 2021a).

Sampling was conducted prior to the major migratory movements involving these species, which occur in March in this area (SEO BirdLife, 2021a). Specifically, mallard faeces (45 faeces) were collected on February 3rd (24 samples) and 11th (21 samples), 2021, while 50 shelduck faeces were collected on February 23rd, 2021. Fresh faeces were collected ensuring that they were firm enough to be handled without disintegration. Each sample was visually inspected to detect any attached sediment. If sediment was observed, it was carefully removed using a clean knife to minimize the risk of contamination from soil propagules. Faeces were carefully collected with a

separation of at least 1 m from each other and stored individually in a paper bag. In the laboratory, samples were dried in open bags at air temperature and stored in dark conditions for three months until the beginning of the experiment. This procedure aimed to prevent predation, fungal growth and sample degradation, and to allow latency periods for those propagules that may need it before hatching (Alekseev et al., 2007).

Hatching experiment design

Thirty faeces of each duck species (60 in total) were randomly selected from those sampled in the field and weighed (ESM Table 1) before the beginning of the experiment. Two temperatures were established, 15 °C (to simulate cold conditions) and 25 °C (warm conditions), with half of the samples (15 faeces of each duck species, 30 in total) allocated to each treatment. A cycle of 12 h light and 12 h darkness was established for all samples, and the light cycle was initiated at the beginning of the inundation. Average light intensity in the chambers was $20 \mu\text{E m}^{-2} \text{s}^{-1}$. The two culture chambers used for the experiment, each programmed at one of the selected temperatures, were BINDER™ KBW 240 (E 5.1) and 720 (E 5.1) (Tuttlingen, Germany), and containers were randomly distributed over the chamber shelves.

The hatching experiment was conducted between May 11th and July 1st, 2021, in plastic containers measuring $10 \times 10 \times 5$ cm (0.5 L). Each container included one faecal sample and was filled with 300 mL of sterilised deionised water. The containers were sealed with a transparent plastic cover to prevent contamination with air droplets between them. Four small holes (1 cm diameter) were drilled in each cover to allow air renewal. Four blank control containers (two in each chamber), with water but without faeces, were added to monitor the possible circulation of propagules in the chambers. The emergence of invertebrates in the containers, including controls, was checked on days 1, 3, 5, 7, 10, 13, 16, 23, 30, 37, and 51 after filling. The procedure consisted of filtering the water through a 20- μm mesh and pouring it carefully. The filtered water was not discarded but was stored temporarily. The 20- μm filter with the faecal material and organisms was washed with deionised water on a Petri dish and examined under a stereomicroscope

at $10\text{--}40\times$ to sort and collect all observed invertebrates. Hatched rotifers were collected with Pasteur pipettes, isolated in plastic tubes, and immediately fixed with 70% ethanol. In order to identify crustaceans to species level (and particularly those with indirect development), emerged juveniles were isolated individually in multiwell plates and placed in the same experimental chamber until reaching adulthood, at which point they were also fixed. During this growth period, copepods and branchiopods were fed every two days with the green algae *Raphidocelis subcapitata* (Korshikov) Nygaard, Komárek, Kristiansen & Skulberg 1987 from a pure laboratory culture; juvenile ostracods were fed with frozen spinach following Schmit et al. (2007). After extracting all observed invertebrate individuals, the Petri dish with the faecal material, the filtered water and the remains of resting propagules captured during the filtration process were returned to their corresponding container to avoid the loss of viable resting propagules. Water loss and evaporation were compensated by adding sterilised deionised water up to the level of 300 mL. The containers were then randomly placed back into the chamber until the next checking period. Water conductivity, oxygen concentration and pH were measured three times (days 6, 16 and 28) in each container (including the controls) during the incubation period with a WTW® Multi 3430 multiparameter probe (with TetraCon 925 for conductivity, FDO 925 for oxygen and WTW® SenTix 940 for pH).

All individuals emerged during the experiment were counted and identified following Koste (1978) for rotifers, Alonso (1996) for branchiopods, Błędzki & Rybak, (2016) and Dussart (1967) for copepods and Meisch (2000) and Karanovic (2012) and references therein for ostracods. Identifications were done at the species level whenever possible. Observed organisms of other taxonomic groups including Ciliophora and Rhabdocoella platyhelminthes (microturbellaria) were also detected and removed (except for ciliates), and their presence was noted during the experiment, although they were not counted or identified to lower taxonomic levels. To identify species as invasive we rely on previous works in the study area in which they discuss the possible origin of the species (Bisquert-Ribes et al., 2023, 2024).

Data analysis

The index for timing of hatching (ITH) was calculated as the average number of days (incubation period) required by each taxon to hatch, as described by Vandekerckhove et al. (2004). This calculation was performed for the entire dataset as well as separately for each duck species and temperature treatment. To assess differences in ITH between temperatures and between ducks, paired t-tests were applied.

To compare the species composition of microinvertebrate communities between temperatures and ducks, we first applied the Hellinger transformation to the species abundance matrix obtained from the experiment in order to reduce the influence of rare and ubiquitous species (Legendre & Gallagher, 2001) using the function *decostand* of the *vegan* R ver. 4.3.1 package (Oksanen et al., 2022; R Core Team, 2023). Then, we obtained a distance matrix with a Bray–Curtis dissimilarity index using the function *vegdist* of the same package. This was used to test for differences in species composition between ducks, temperature treatments and along the experiment by means of PERMANOVA with the function *adonis2* of the *vegan* package.

Finally, to assess the main patterns in the ordination of the microinvertebrate assemblages for the four datasets (two ducks x two temperatures), we performed a nonmetric multidimensional scaling analysis (NMDS) using the function *metaMDS* of the *vegan* package.

Results

A total of 34 species were identified from the experiment, including rotifers and crustaceans (cladocerans, copepods, and ostracods; Table 1). More than twice as many species (26) were detected in the 25 °C treatment (warm conditions) compared to the colder (15 °C) temperature treatment (12 species). Out of the 15 rotifer species, 13 hatched exclusively at the higher temperature, with none hatching at both temperatures. Among the 19 crustacean species identified, nine (eight ostracods and one cladoceran) hatched only at 25 °C, six (four ostracods, one cladoceran, and one copepod) hatched only at 15 °C, and four ostracods hatched at both temperatures (Table 1). When comparing the two duck species, 30 different

invertebrates were recorded in the shelduck samples, whereas only 11 were detected in the mallard samples. In the combined treatment analysis (temperature x duck), more species were found in the shelduck samples (11 and 23 species at cold and warm conditions, respectively) than in the mallard samples (three and nine species, respectively). The most frequently occurring species were the ostracods *Ilyocypris gibba*, *Heterocypris incongruens*, and the rotifer *Lecane tenuiseta*, each detected in nine different samples, followed by *Heterocypris* sp. 3, which appeared in six samples. Among the detected species, five invasive ostracods were found (Table 1), appearing exclusively in shelduck samples and at the 25 °C treatment, except for *Sarscypridopsis* sp. B (ostracod), which appeared in both temperature treatments. Individuals belonging to Rhabdocoela were observed in one shelduck sample at 25 °C. Members of Ciliophora were present in 10 different samples of the experiment including both temperatures and duck species. A significant percentage of the samples, 52% (31 out of 60 samples), showed no hatching throughout the experiment. When comparing the two temperatures, the proportion of non-hatching samples was considerably higher at 15 °C, with 80% of the samples (24 out of 30 samples) failing to hatch, while at 25 °C, only 23% of the samples (seven out of 30 samples) did not hatch. In terms of duck species, over half of the samples showed no hatching in both cases, with 50% in mallards and 53% in shelducks.

Since hatchlings began to emerge, the accumulated species richness for shelduck was consistently higher than that for mallards across both temperature treatments (Fig. 1). In the warmer treatment, no individuals appeared before the 7th day of the experiment. Particularly noteworthy was the shelduck treatment at 25 °C, which accumulated species richness did not appear to stabilize throughout the experiment. In contrast, mallard samples at 25 °C exhibited fewer species and showed two stabilization periods: one at the beginning and another between days 30 and 40 of the experiment, with a slight increase in new species towards the end. In the cold treatment, hatchlings took up to 30 days to appear in both duck species. Shelduck at 15 °C exhibited species accumulation values similar to those of mallard at 25 °C, ultimately surpassing it in species count by the end of the experiment. As with the other temperature treatment, mallard showed fewer species than shelduck

Table 1 Index for timing of hatching (ITH) values for invertebrate species hatched from faecal samples in the experiment, separated by duck species (mallard and shelduck) and temperature treatment (15 °C and 25 °C)

Species list	Species code	ITH (days)				
		Total	Mallard	Shelduck	15 °C	25 °C
Rotifera						
<i>Brachionus calyciflorus</i> Pallas 1766	Bra caly	13 [†]		13 [†]		13 [†]
<i>Cephalodella</i> cf. <i>obvia</i> Donner 1950 (FR)	Cep obvi	33	33		33	
<i>Cephalodella gibba</i> (Ehrenberg 1838) (FR)	Cep gibb	32	38	29		32
<i>Cephalodella</i> sp. 1 (FR)	Cep sp1	51		51		51
<i>Collotheca</i> sp. (FR)	Col sp	51 [†]		51 [†]		51 [†]
<i>Encentrum</i> cf. <i>longipes</i> Wulfert 1936 (FR)	Enc long	37		37		37
<i>Itura</i> cf. <i>aurita</i> (Ehrenberg 1830) (FR)	Itu auri	51		51	51	
<i>Lecane bulla</i> (Gosse 1851) (FR)	Lec bull	51		51		51
<i>Lecane</i> cf. <i>hornemanni</i> (Ehrenberg 1834) (FR)	Lec horn	51	51			51
<i>Lecane closteroerca</i> (Schmarda 1859) (FR)	Lec clos	51 [†]		51 [†]		51 [†]
<i>Lecane tenuiseta</i> Harring 1914 (FR)	Lec tenu	42	47	42		42
<i>Lepadella patella</i> (O. F. Müller 1773) (FR)	Lep pate	43		43		43
<i>Proales</i> cf. <i>similis</i> de Beauchamp 1907 (FR)	Pro simi	51 [†]	52 [†]			51 [†]
<i>Proales</i> cf. <i>syltensis</i> Tzschaschel 1979 (FR)	Pro sylt	51	51	51		51
<i>Trichocerca</i> cf. <i>cavia</i> (Gosse 1886) (FR)	Tri cavi	41	41			41
Branchiopoda						
<i>Daphnia magna</i> Straus 1820	Dap magn	37 [†]		37 [†]	37 [†]	
<i>Macrothrix laticornis</i> (Jurine 1820)	Mac lati	13 [†]		13 [†]		13 [†]
Copepoda						
Calanoida	Calan	51 [†]		51 [†]	51 [†]	
Ostracoda						
<i>Cypretta</i> cf. <i>maya</i> Cohuo-Durán et al. 2013 * (FR)	Cyp maya	30		30		30
<i>Cypridopsis vidua</i> (Müller 1776)	Cyp vidu	49		49		49
<i>Cypris decaryi</i> Gauthier 1933 * (FR)	Cyp deca	30 [†]		30 [†]		30 [†]
<i>Eucypris virens</i> (Jurine 1820)	Euc vire	47	44	51	47	
<i>Hemicypris exigua</i> Broodbakker 1983 * (FR)	Hem exig	45		45		45
<i>Heterocypris incongruens</i> (Ramdohr 1808)	Het inco	40	41	39	41	40
<i>Heterocypris salina</i> (Brady 1868)	Het sali	13 [†]		13 [†]		13 [†]
<i>Heterocypris</i> sp. 1 (FR)	Het sp1	27		27		27
<i>Heterocypris</i> sp. 3 (FR)	Het sp3	37	36	37	39	37
<i>Ilyocypris biplicata</i> var. <i>anomala</i> (Koch 1838) (FR)	Ily bipl	51 [†]		51 [†]	51 [†]	
<i>Ilyocypris getica</i> Masi 1906 (FR)	Ily geti	40		40		40
<i>Ilyocypris gibba</i> (Ramdohr 1808)	Ily gibb	39	30	39	43	38
<i>Isocypris beauchampi</i> (Paris 1920) * (FR)	Iso beau	16 [†]		16 [†]		16 [†]
<i>Sarscypridopsis aculeata</i> (Costa 1847)	Sar acul	39		39	39	
<i>Sarscypridopsis</i> sp. B * (FR)	Sar spB	41		41	40	41
<i>Trajancypris clavata</i> (Baird 1838) (FR)	Tra clav	51		51	51	
Mean ± SD		39.5 ± 12.0	42.1 ± 7.4	38.9 ± 12.6	43.6 ± 6.4	37.8 ± 12.7

Total ITH was calculated using all individuals of the same species that hatched in the experiment. ITH values with the symbol † correspond to species with only one individual hatched, so ITH does not represent mean but unique values. An asterisk (*) next to species names indicates the species is invasive. Species not detected in previous zoochory studies have been indicated as first record (FR) next to the species name

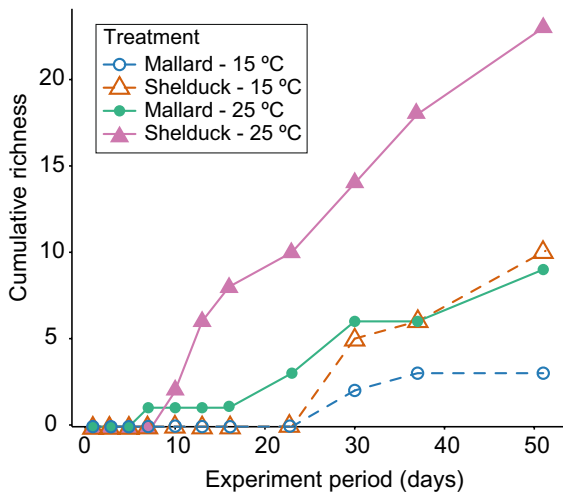


Fig. 1 Cumulative species richness for each treatment (duck species x temperature) along the hatching experiment

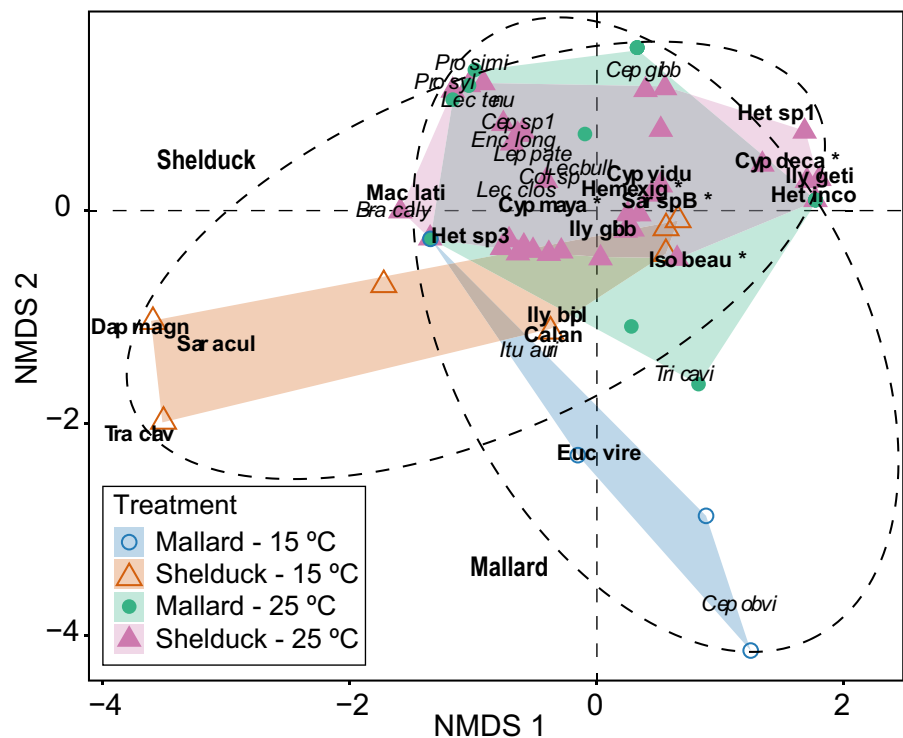
in cold conditions. Regarding water characteristics along the experiment, when comparing both temperature treatments, we found very similar values of water conductivity ($318 \pm 106 \mu\text{S}/\text{cm}$ and $353 \pm 130 \mu\text{S}/\text{cm}$, respectively) and pH (7.6 ± 0.5 and 8.0 ± 0.6 , respectively). Oxygen saturation in the cold treatment of the

experiment was $69 \pm 25\%$, slightly lower than in the warm treatment ($80 \pm 16\%$).

The ITH indicated that species took an average of approximately 40 days to hatch (Table 1). At 15 °C, the average hatching time was delayed to 44 days, whereas at 25 °C it was about six days earlier. However, this difference was not statistically significant ($t=1.46$; d.f.=3; $p=0.24$). Similarly, when comparing between duck species, the average ITH was about 42 days for mallard and 39 days for shelduck, but again, the difference was not significant ($t=-0.06$; d.f.=6; $p=0.95$).

The PERMANOVA analysis revealed that the invertebrate species composition varied between temperatures ($F=2.49$; $p=0.01$). This variation was evident in the NMDS ordination (Fig. 2), which separated samples along the second axis in relation to experimental temperature: most warm samples were positioned on the positive side of the axis (with some on the negative side), while cold samples were ordered on the negative side. Additionally, PERMANOVA indicated differences in species composition between duck species ($F=5.16$; $p=0.01$), throughout the experiment ($F=1.65$; $p<0.01$), and in the interaction between temperature and duck

Fig. 2 Non-metric multidimensional scaling (NMDS) ordination of the experiment samples and invertebrate species hatched, according to duck species and temperature. The shaded areas encompassing the samples (convex hull) correspond to each treatment. An asterisk (*) next to the species code indicates the species is invasive, species codes in *italics* indicate rotifers and **bold** type indicates crustaceans. See Table 1 for species codes



species ($F=1.96$; $p=0.01$). The NMDS ordination also showed a clear differentiation between the two duck species at 15 °C, likely due to the emergence of different species, but not so much at 25 °C. Under cold conditions, the crustaceans *Trajancypis clavata* (ostracod) and *Daphnia magna* (branchiopod) were dominant in shelduck samples, whereas the ostracod *Eucypris virens* and the rotifer *Cephalodella* cf. *obvia* were prevalent in mallard samples. In warm samples, the NMDS showed a similar and overlapping dispersion between the two species. Shelduck samples were widely dispersed across the area, while mallard samples were more concentrated around the outer edges. This distribution likely contributed to the differences in species composition between the two temperature treatments for mallard, as indicated by the PERMANOVA pairwise test analysis (ESM Table 2). The analysis also revealed significant differences in most other comparisons, except between the two temperature treatments for shelduck. This exception might be due to three shelduck samples at 15 °C having a species composition very similar to some samples at 25 °C, contrasting with the other four samples, which were more distinct (Fig. 2).

Discussion

This study aimed to assess the role of two waterfowl species on the dispersal through endozoochory of aquatic invertebrates in a Mediterranean wetland, with a focus on how temperature influences hatching success and community composition. We found higher species richness and hatching rates at warmer conditions, and shelducks showed a more diverse invertebrate assemblage than mallards, including several invasive ostracods. We recorded 34 taxa belonging to a broad array of organisms: rotifers, ostracods, cladocerans, copepods, ciliates, and rhabdocoelans. Several rotifers and ostracods we detected (e.g., *Collothea* sp., *Lepadella patella*, *Isocypris beauchampi*, and *T. clavata*, see Table 1), so as microturbellarians, represent, to our knowledge, the first records of internal transport by waterbirds. We found five invasive ostracod species in our samples, including *Hemicypris exigua*, *Cypris decaryi* and *Sarscypridopsis* sp. B, which, as far as we know, have not previously been reported in endozoochory studies, although bird-dispersal of species of the same genera

has been documented elsewhere (Proctor & Malone, 1965; Proctor et al., 1967; Valls et al., 2017; Lovas-Kiss et al., 2018). Our findings, along with previous studies (Lovas-Kiss et al., 2018; Martín-Vélez et al., 2022), illustrate that passively dispersing species, irrespective of their origin, benefit from waterfowl endozoochory, which can have an influence on ecosystems at local, regional, and continental scales (Green et al., 2002; Reynolds et al., 2015; Green, 2016).

A few studies have documented the survival of copepods after waterbird gut passage. Frisch et al. (2007) and Lovas-Kiss et al. (2018) reported the presence of harpacticoids and cyclopoids, while Valls et al. (2017), similarly to our findings, reported an unidentified calanoid species. However, in all these studies, other groups were more abundant than copepods. The apparently low importance of internal dispersal of copepods by waterbirds may be related to the fact that most copepod species do not produce resting eggs (Alekseev et al., 2007; Błędzki & Rybak, 2016). Instead, many of them enter dormancy at the copepodite stage, a developmental instar that may be more susceptible to digestive degradation during gut passage than more resistant resting eggs, potentially limiting their capacity for successful endozoochory dispersal. Although not the primary focus of this study, the detection of Rhabdocoela and Ciliophora in several samples is notable. While ciliates have been reported before (Valls et al., 2017; Silva et al., 2021), the presence of Rhabdocoela, as far as we know, has not been reported in previous endozoochory studies, and their occurrence here can suggest that they may also be capable of passive dispersal via waterbirds. A previous work by Rogers (2014) mentions the culture of turbellarian cocoons, but he did not specify if these cocoons were obtained from bird faces or gastrointestinal samples, or either from mud collected from pools, boots or vehicle. Further research could explore the potential dispersal dynamics of these groups by endozoochory. Ducks are also likely to disperse other groups of organisms whose propagules do not hatch easily under laboratory conditions, such as plumatellid bryozoans (Martín-Vélez et al., 2022).

Comparing temperature treatments, more species hatched at the warmer treatment, and they did it earlier on average. The species that hatched at both temperatures were different as well. Some species that typically appear in winter conditions in the region,

such as *D. magna*, *S. aculeata*, *T. clavata*, and *E. virens* (Bisquert-Ribes et al., 2024), hatched only at the lower temperature tested. Conversely, many species typical of summer conditions in the area appeared at the higher temperature, including the branchiopod *Macrothrix laticornis*, and the invasive ostracods *C. decaryi*, *Cyprretta* cf. *maya*, and *H. exigua*, (Bisquert-Ribes et al., 2024). In this previous work in the study area, all crustacean species reported in the present work were detected except for *Heterocypris* sp. 1, *T. clavata* and *I. biplicata* var. *anomala*. However, the first two species were detected in another study in the rice fields of the Albufera (Bisquert-Ribes et al., 2023). This may suggest that temperature can influence the hatching success and species composition, as it has been reported as a highly relevant factor for the emergence of resting propagules (Alekseev et al., 2007). Indeed, temperature changes can trigger specific life-cycle phases in aquatic organisms, such as the development or hatching of diapausing eggs (Alekseev et al., 2007; Bonacina et al., 2023).

In our study, we focused exclusively on temperature as a factor influencing hatching success, however, other factors can be determinant too. For example, latency period can be relevant, as many resting propagules require relatively long latency periods before hatching (Alekseev et al., 2007). In this study, more species typical of summer temperatures hatched, perhaps because their resting eggs were present since the previous summer, so they may have had a longer latency period that allowed them to quickly respond to inundation triggering hatching. In contrast, most resting propagules of typical wintering species (which would probably hatch at lower temperatures) may have been produced during the winter of the same year in which the samples were collected, and the experiment was conducted, and therefore were younger. Thus, it is possible that species requiring longer latency periods did not hatch. Further research is needed to verify this hypothesis.

Other environmental factors such as salinity (Valls et al., 2017; Conde-Porcuna et al., 2018), photoperiod, or even the interaction between different factors may also have appreciable effects in the hatching success (Conde-Porcuna et al., 2018). In this sense, unlike other studies that isolate resting eggs before incubation (e.g., Brochet et al., 2010; Martín-Vélez et al., 2022), we used entire faecal samples, which may have altered water chemistry within experimental

containers. For instance, organic matter and microbial or algal growth could have influenced oxygen levels, potentially masking or modifying temperature effects. This could be the case of the unexpected lower oxygen levels detected in the colder treatment of this study. Possibly, a higher concentration of heterotrophic bacteria in some samples of the cold treatment or algal activity in the warmer samples may have increased respiration or photosynthesis, respectively, leading to differences in oxygen content. These methodological aspects should be considered when interpreting our results and suggest the need for more controlled approaches and for testing the interaction among multiple environmental factors affecting propagule hatching (Spencer & Blaustein, 2001; Alekseev et al., 2007). Nevertheless, it should be noticed that the eggs of some aquatic invertebrates, such as ostracods, are deposited attached to physical or organic structures in the sediment (e.g., sand grains, shells, leaves...) (Aguilar-Alberola & Mesquita-Joanes, 2013), and therefore standard methods used to separate eggs through density centrifugation may not be appropriate to uncover all the diversity of propagules in a given sample.

Gut passage and retention time (García-Álvarez et al., 2015), bird diet (Clark, 2021), or propagule traits such as size (Santamaría et al., 2023) can also be determinant for the hatching success of invertebrate propagules. Some studies have shown that the shape and volume of propagules can provide an advantage to invaders over native species (García-Álvarez et al., 2015; Lovas-Kiss et al., 2023). While invasive species may experience less propagule pressure, favourable environmental characteristics or conditions, or even the characteristics of their propagules, may facilitate their establishment.

Another important consideration is the likely underestimation of the number of taxa effectively dispersed by wintering waterfowl in the Albufera. Our sampling represents only a small fraction of the approximately 20,000 ducks that arrive each winter to the area (SEO BirdLife, 2021a). Moreover, although ducks have been documented as a very relevant group for the efficient endozoochoric dispersal of aquatic invertebrates (Figuerola et al., 2003; Brochet et al., 2010; Valls et al., 2017; Moreno et al., 2019), other bird groups such as gulls and shorebirds, are also known to disperse aquatic invertebrate propagules (Conde-Porcuna et al., 2018; Lovas-Kiss et al., 2018;

Martín-Vélez et al., 2022), and consequently the great variety of invertebrates being dispersed by birds are most probably underestimated in our study.

Our results suggest that both duck species, shelduck and mallard, can serve as effective dispersers of invertebrate propagules, consistent with previous studies (Valls et al., 2017; Green et al., 2023). However, shelduck samples showed a higher diversity of invertebrates compared to mallard samples. Mallard and shelduck are both plastic species that adapt their foraging methods according to several environmental variables, such as habitat and prey availability (Ferns & Reed, 2009; Viain et al., 2011; Bezzalla et al., 2019). Their feeding behaviour involves techniques like upending, head dipping, and sieving sediments (Liordos, 2010) that may have favoured the ingestion of the studied species. The abundance of rotifer genera typically found in the littoral (*Lecane*, *Cephalodella*, *Lepadella*) or in the benthos (*Encentrum*, *Itura*, *Proales*, see Koste, 1978), also supports this feeding behaviour. Regarding niche and foraging habitat preferences, both species overlap during winter, foraging in flooded rice fields in the area, although shelducks often prefer saline and deeper environments for roosting (SEO BirdLife, 2021a). This aligns with the detection of the ostracod *Heterocypris salina*, a common species in halophilic environments (Meisch, 2000), particularly in the area (Rueda et al., 2006). Variations in the egg bank composition of the sediments from which the birds feed could also explain the differences in species composition detected between the two waterbirds. Many invertebrate species have patchy distributions, so their propagule egg banks are heterogeneous (Brendonck & De Meester, 2003; Pinel-Alloul & Ghadouani, 2007), affecting the diversity of resting propagules ingested by the birds. In any case, this study highlights the strong potential of both duck species to disperse propagules from a wide range of invertebrate groups, including rotifers, crustaceans, ciliates, and flatworms. However, previous studies have shown that dispersal by these birds is not limited to resting propagules; they are also capable of dispersing organisms such as aquatic beetles (Laux & Kölsch, 2014) and snails (Cadée, 2011; van Leeuwen et al., 2012).

Given the ability of these birds to effectively disperse a wide array of invertebrates, their movements across regions could be determinant in the spread of invasive species, which this study shows they are

capable of dispersing. Mallard is the most abundant Anatidae in the Albufera in winter, and a relevant portion of the population remains in the region to breed (SEO BirdLife, 2021a; b). As a result, its potential for long-distance dispersal to other regions may be somewhat reduced. Nevertheless, mallard are also highly abundant throughout Europe (Drilling et al., 2020), and their seasonal migrations, southward to warmer regions for overwintering and northward for breeding (Díaz et al., 1996; SEO BirdLife, 2012), make it a strong candidate for interregional dispersal of aquatic organisms, including invasive ones. Shelduck has experienced a positive population growth trend in Europe (Fouque et al., 2009; Martínez-Abraín et al., 2016; Nagy & Langendoen, 2020), including the wintering population in the Albufera, which has increased significantly to exceed 2,000 birds in last years (SEO BirdLife, 2021a). This population growth, along with its generalist habitat preferences with selection for artificial wetlands such as rice fields (Walmsley & Moser, 1981; Rendón et al., 2008), as well as its seasonal movements (Cimiotti et al., 2024) and migratory patterns between populations in central and southern Europe (Cimiotti et al., 2025), position the common shelduck as a potential vector for the spread of invasive species. This is particularly relevant in our study area, where previous research has documented a high diversity of invasive ostracods in ricefields (Bisquert-Ribes et al., 2023, 2024). In a global change scenario, increasing mobility and range expansion of waterbirds (Pavón-Jordán et al., 2015; Ramírez et al., 2018) may further facilitate the spread of such invasive species across a variety of European wetlands.

Conclusions

This study provides new evidence that mallards and shelducks can disperse a taxonomically diverse assemblage of aquatic invertebrates via endozoochory, including several invasive ostracod species and other groups previously unreported, such as flatworms. By testing the effect of temperature on hatching success, we show that warmer conditions may favour the emergence of more species, including invaders, and reveal differences in the dispersal capacity between the two bird species. We acknowledge the importance of considering other

environmental factors, such as photoperiod, conductivity, pH, and their potential interactions, as they can also significantly influence the hatching success of many propagules (Alekseev et al., 2007). Including these variables in future studies will help improve our understanding of how bird-dispersed resting propagules respond to environmental conditions and how the resulting species may impact local aquatic communities.

Acknowledgements We would like to thank the technical staff at the Cavanilles Institute for their assistance in measuring container conditions throughout the experiment. We are very grateful to Andy Green and one anonymous reviewer for their helpful suggestions on a previous version of the manuscript.

Funding This work was supported by Comunitat Valenciana Department of Innovation, Universities, Science and Digital Society [EXOCRUST AICO/2020/182, 2020] and Spanish Ministry of Education and Vocational Training [FPU19/02264, 2020]. Partial support also provided by project PID2023-146630NB-I00 funded by MICIU/AEI/10.13039/501100011033 and by FEDER, EU.

Data availability Raw data and R scripts of this study are available at Zenodo (<https://doi.org/10.5281/zenodo.14640283>).

Declarations

Conflict of interest The authors have no competing interests to declare that are relevant to the content of this article.

Ethical approval No approval of research ethics committees was required to accomplish the goals of this study because work was conducted with chitinized remains.

Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>.

References

- Aguilar-Alberola, J. A. & F. Mesquita-Joanes, 2013. The ontogeny of *Heterocypris bosniaca* (Ostracoda: Cyprididae): description of postembryonic instars and rediscovery of the neglected A-9 stage. *Journal of Crustacean Biology* 33: 348–371. <https://doi.org/10.1163/1937240X-00002133>.
- Alekseev, V. R., B. T. de Stasio & J. J. Gilbert, 2007. *Diapause in Aquatic Invertebrates: Theory and Human Use*, Springer, Dordrecht.
- Alonso, M., 1996. Crustacea: Branchiopoda, Fauna Ibérica, Consejo Superior de Investigaciones Científicas (CSIC).
- Bezzalla, A., M. Houhamdi, M. Cherif Maazi & H. Chenchouni, 2019. Modelling climate influences on population dynamics and diurnal time budget of the Shelduck (*Tadorna tadorna*) wintering in Ramsar wetlands of Algeria. *Avian Biology Research* 12: 77–95. <https://doi.org/10.1177/1758155919835122>.
- BirdLife, S. E. O., 2012. Atlas de las aves en invierno en España 2007–2010, Ministerio de Agricultura, Alimentación y Medio Ambiente-SEO BirdLife, Madrid.
- Bisquert-Ribes, M., E. García-Berthou, M. A. Redón-Morte, J. Rueda, F. Mesquita-Joanes & X. Armengol, 2024. Are rice fields less diverse and more invaded by non-native species than less impacted habitats? A test with wetland microcrustaceans. *Agriculture, Ecosystems & Environment* 378: 109305. <https://doi.org/10.1016/j.agee.2024.109305>.
- Bisquert-Ribes, M., D. J. Horne, J. M. Benavent, R. Martínez, P. Vera, J. Rueda & F. Mesquita-Joanes, 2023. High incidence of exotic ostracods in the rice fields of a protected Mediterranean wetland. *Inland Waters* 13: 428–445. <https://doi.org/10.1080/20442041.2023.2262353>.
- Błędzki, L. A. & J. I. Rybak, 2016. Freshwater Crustacean Zooplankton of Europe, Springer, Switzerland.
- Bonacina, L., F. Fasano, V. Mezzanotte & R. Fornaroli, 2023. Effects of water temperature on freshwater macroinvertebrates: a systematic review. *Biological Reviews* 98: 191–221. <https://doi.org/10.1111/BRV.12903>.
- Brendonck, L. & L. De Meester, 2003. Egg banks in freshwater zooplankton: evolutionary and ecological archives in the sediment. *Hydrobiologia* 491: 65–84. <https://doi.org/10.1023/A:1024454905119>.
- Brendonck, L., M. Jocque, A. Hulsmans & B. Vanschoenwinkel, 2010. Pools “on the rocks”: freshwater rock pools as model system in ecological and evolutionary research. *Limnetica* 29: 25–40. <https://doi.org/10.23818/limn.29.03>.
- Brochet, A. L., M. Gauthier-Clerc, M. Guillemain, H. Fritz, A. Waterkeyn, Á. Baltanás & A. J. Green, 2010. Field evidence of dispersal of branchiopods, ostracods and bryozoans by teal (*Anas crecca*) in the Camargue (southern France). *Hydrobiologia* 637: 255–261. <https://doi.org/10.1007/s10750-009-9975-6>.
- Cadée, G. C., 2011. *Hydrobia* as “Jonah in the whale”: shell repair after passing through the digestive tract of shelducks alive. *Palaos* 26: 245–249. <https://doi.org/10.2110/palo.2010.p10-095r>.
- Cimiotti, D. S., D. V. Cimiotti, H. Hötter & S. Garthe, 2025. Ringing, tracking and counting data reveal five wintering patterns in European Common Shelducks. *Ibis* 167: 5–24. <https://doi.org/10.1111/IBI.13278>.
- Cimiotti, D. S., H. Hötter & S. Garthe, 2024. Exploratory and seasonal movements of adult common shelducks in the

- eastern Wadden Sea. *Journal of Ornithology* 165: 289–300. <https://doi.org/10.1007/S10336-023-02128-X>.
- Clark, D., 2021. Assessing dietary factors that influence branchiopod cyst viability rates following mallard digestion and excretion. Honors Projects. 664.
- Conde-Porcuna, J. M., C. Pérez-Martínez & E. Moreno, 2018. Variations in the hatching response of rotifers to salinity and waterbird ingestion. *Journal of Plankton Research* 40: 326–341. <https://doi.org/10.1093/PLANKT/FBY010>.
- Díaz, M., B. Asensio, & J. L. Tellería, 1996. Aves ibéricas I. No passeriformes. J. M. Reyero Editor. Madrid.
- Drilling, N., R. D. Titman, & F. McKinney, 2020. Mallard (*Anas platyrhynchos*), version 1.0. In *Birds of the World* (S. M. Billerman, Editor). Cornell Lab of Ornithology, Ithaca, NY, USA. <https://doi.org/10.2173/bow.mallara3.01>.
- Dussart, B., 1967. Les copépodes des eaux continentales d'Europe occidentale. Tome I: Calanoides et Harpacticoides. N. Boubée & Cie, Paris, France.
- Ferns, P. N. & J. P. Reed, 2009. Effects of the Cardiff Bay tidal barrage on the abundance, ecology and behaviour of shelducks *Tadorna tadorna*. *Aquatic Conservation: Marine and Freshwater Ecosystems* 19: 466–473. <https://doi.org/10.1002/AQC.1011>.
- Figuerola, J. & A. J. Green, 2002. Dispersal of aquatic organisms by waterbirds: a review of past research and priorities for future studies. *Freshwater Biology* 47: 483–494. <https://doi.org/10.1046/J.1365-2427.2002.00829.X>.
- Figuerola, J., A. J. Green & L. Santamaría, 2003. Passive internal transport of aquatic organisms by waterfowl in Doñana, south-west Spain. *Global Ecology and Biogeography* 12: 427–436. <https://doi.org/10.1046/J.1466-822X.2003.00043.X>.
- Fouque, C., M. Guilleman & V. Schricke, 2009. Trends in the numbers of Coot *Fulica atra* and wildfowl Anatidae wintering in France, and their relationship with hunting activity at wetland sites. *Wildfowl Special Issue* 2: 42–59.
- Frisch, D., A. J. Green & J. Figuerola, 2007. High dispersal capacity of a broad spectrum of aquatic invertebrates via waterbirds. *Aquatic Sciences* 69: 568–574. <https://doi.org/10.1007/s00027-007-0915-0>.
- Gálvez, Á., A. E. Magurran, X. Armengol, S. Savatnalinton & F. Mesquita-Joanes, 2022. Metacommunity structure and dynamics. In Dalu, T. & R. J. Wasserman (eds), *Fundamentals of Tropical Freshwater Wetlands: From Ecology to Conservation Management* Elsevier: 549–586. <https://doi.org/10.1016/B978-0-12-822362-8.00011-6>.
- García-Álvarez, A., C. H. A. van Leeuwen, C. J. Luque, A. Hussner, A. Vélez-Martín, A. Pérez-Vázquez, A. J. Green & E. M. Castellanos, 2015. Internal transport of alien and native plants by geese and ducks: an experimental study. *Freshwater Biology* 60: 1316–1329. <https://doi.org/10.1111/FWB.12567>.
- Green, A. J., 2016. The importance of waterbirds as an overlooked pathway of invasion for alien species. *Diversity and Distributions* 22: 239–247. <https://doi.org/10.1111/DDI.12392>.
- Green, A. J. & J. Elmberg, 2014. Ecosystem services provided by waterbirds. *Biological Reviews* 89: 105–122. <https://doi.org/10.1111/BRV.12045>.
- Green, A. J., J. Figuerola & M. I. Sánchez, 2002. Implications of waterbird ecology for the dispersal of aquatic organisms. *Acta Oecologica* 23: 177–189. [https://doi.org/10.1016/S1146-609X\(02\)01149-9](https://doi.org/10.1016/S1146-609X(02)01149-9).
- Green, A. J., K. M. Jenkins, D. Bell, P. J. Morris & R. T. Kingsford, 2008. The potential role of waterbirds in dispersing invertebrates and plants in arid Australia. *Freshwater Biology* 53: 380–392. <https://doi.org/10.1111/J.1365-2427.2007.01901.X>.
- Green, A. J., Á. Lovas-Kiss, C. Reynolds, E. Sebastián-González, G. G. Silva, C. H. A. van Leeuwen & D. M. Wilkinson, 2023. Dispersal of aquatic and terrestrial organisms by waterbirds: A review of current knowledge and future priorities. *Freshwater Biology* 68: 173–190. <https://doi.org/10.1111/fwb.14038>.
- Green, A. J., M. Soons, A. L. Brochet & E. Kleyheeg, 2016. Dispersal of plants by waterbirds. Chapter six. In Sekercioglu, Ç. H., D. G. Wenny & C. J. Whelan (eds), *Why Birds Matter: Avian Ecological Function and Ecosystem Services* University of Chicago Press: 107–145.
- Heino, J., A. S. Melo, T. Siqueira, J. Soininen, S. Valanko & L. M. Bini, 2015. Metacommunity organisation, spatial extent and dispersal in aquatic systems: patterns, processes and prospects. *Freshwater Biology* 60: 845–869. <https://doi.org/10.1111/FWB.12533>.
- Hulsmans, A., K. Moreau, L. De Meester, B. J. Riddoch & L. Brendonck, 2007. Direct and indirect measures of dispersal in the fairy shrimp *Branchipodopsis wolffi* indicate a small scale isolation-by-distance pattern. *Limnology and Oceanography* 52: 676–684. <https://doi.org/10.4319/LO.2007.52.2.0676>.
- Karanovic, I., 2012. Recent freshwater ostracods of the world. Crustacea, Ostracoda, Podocopida, Springer, Berlin, Heidelberg.
- Kleyheeg, E., M. Klaassen & M. B. Soons, 2016. Seed dispersal potential by wild mallard duck as estimated from digestive tract analysis. *Freshwater Biology* 61: 1746–1758. <https://doi.org/10.1111/FWB.12814>.
- Koste, W., 1978. Rotatoria die rädertiere mitteleuropas. Monogononta. Gebrüder Brontraeger, Berlin, Germany.
- Laux, J.-J. & G. Kölsch, 2014. Potential for passive internal dispersal: eggs of an aquatic leaf beetle survive passage through the digestive system of mallards. *Ecological Entomology* 39: 391–394. <https://doi.org/10.1111/EEN.12097>.
- Legendre, P. & E. D. Gallagher, 2001. Ecologically meaningful transformations for ordination of species data. *Oecologia* 129: 271–280. <https://doi.org/10.1007/s004420100716>.
- Leibold, M. A. & J. M. Chase, 2018. *Metacommunity Ecology*, Princeton University Press.
- Leibold, M. A., M. Holyoak, N. Mouquet, P. Amarasekare, J. M. Chase, M. F. Hoopes, R. D. Holt, J. B. Shurin, R. Law, D. Tilman, M. Loreau & A. Gonzalez, 2004. The metacommunity concept: a framework for multi-scale community ecology. *Ecology Letters* 7: 601–613. <https://doi.org/10.1111/J.1461-0248.2004.00608.X>.
- Liordos, V., 2010. Foraging guilds of waterbirds wintering in a Mediterranean coastal wetland. *Zoological Studies* 49: 311–323.
- Lovas-Kiss, Á., M. J. Navarro-Ramos, O. Vincze, V. Löki, R. Ugyán, F. Pallér-Kapusi, C. H. A. van Leeuwen, A. J. Green & B. A. Lukács, 2023. Traits for transport: alien

- wetland plants gain an advantage during endozoochorous seed dispersal by waterfowl. *Freshwater Biology* 68: 1703–1715. <https://doi.org/10.1111/FWB.14154>.
- Lovas-Kiss, Á., M. I. Sánchez, A. V. Molnár, L. Valls, X. Armengol, F. Mesquita-Joanes & A. J. Green, 2018. Crayfish invasion facilitates dispersal of plants and invertebrates by gulls. *Freshwater Biology* 63: 392–404. <https://doi.org/10.1111/FWB.13080>.
- Lovas-Kiss, Á., M. I. Sánchez, D. M. Wilkinson, N. E. Coughlan, J. A. Alves & A. J. Green, 2019. Shorebirds as important vectors for plant dispersal in Europe. *Ecography* 42: 956–967. <https://doi.org/10.1111/ECOG.04065>.
- Martínez-Abraín, A., J. Jiménez, J. A. Gómez & D. Oro, 2016. Differential waterbird population dynamics after long-term protection: the influence of diet and habitat type. *Spanish Society of Ornithology. Ardeola* 63: 79–101. <https://doi.org/10.13157/ARLA.63.1.2016.RP4>.
- Martín-Vélez, V., M. I. Sánchez, Á. Lovas-Kiss, F. Hortas & A. J. Green, 2022. Dispersal of aquatic invertebrates by lesser black-backed gulls and white storks within and between inland habitats. *Aquatic Sciences* 84: 10. <https://doi.org/10.1007/s00027-021-00842-3>.
- McKenzie, K. G. & A. Moroni, 1986. Man as an agent of crustacean passive dispersal via useful plants - exemplified by *Ostracoda ospiti esteri* of the Italian ricefields ecosystem - and Implications arising therefrom. *Journal of Crustacean Biology* 6: 181–198.
- Meisch, C., 2000. *Freshwater Ostracoda of Western and Central Europe*. In Schwoerbel, J., & P. Zwick (eds) *Süßwasserfauna von Mitteleuropa*. Spektrum Akademischer Verlag. pp. 1–522.
- Moreno, E., C. Pérez-Martínez & J. M. Conde-Porcuna, 2019. Dispersal of rotifers and cladocerans by waterbirds: seasonal changes and hatching success. *Hydrobiologia* 834: 145–162. <https://doi.org/10.1007/S10750-019-3919-6>.
- Nagy, S. & T. Langendoen, 2020. Flyway trend analyses based on data from the African Eurasian Waterbird Census from the period of 1967–2018, Online publication, Wageningen, The Netherlands.
- Oksanen, J., G. L. Simpson, F. G. Blanchet, R. Kindt, P. Legendre, P. R. Minchin, R. B. O'Hara, P. Solymos, M. H. H. Stevens, E. Szoecs, H. Wagner, M. Barbour, M. Bedward, B. Bolker, D. Borcard, G. Carvalho, M. Chirico, M. De Caceres, S. Durand, H. B. A. Evangelista, R. FitzJohn, M. Friendly, B. Furneaux, G. Hannigan, M. O. Hill, L. Lahti, D. McGlinn, M.-H. Ouellette, E. Ribeiro Cunha, T. Smith, A. Stier, C. J. F. Ter Braak, & J. Weedon, 2022. *vegan: community ecology package: ordination, diversity and dissimilarities*. R Package version 2.6–4. [accessed 2024 Nov 3]. <https://Cran.R-project.org/package=vegan>.
- Olmo, C., M. Antón-Pardo, R. Ortells & X. Armengol, 2020. Influence of restoration age on egg bank richness and composition: an ex situ experiment. *Journal of Plankton Research* 42: 553–563. <https://doi.org/10.1093/plankt/fbaa034>.
- Patterson, I. J., 1982. *The shelduck*, The University Press, Cambridge, England.
- Pavón-Jordán, D., A. D. Fox, P. Clausen, M. Dagys, B. Deceuninck, K. Devos, R. D. Hearn, C. A. Holt, M. Hornman, V. Keller, T. Langendoen, L. Lawicki, S. H. Lorentsen, L. Luigujõe, W. Meissner, P. Musil, L. Nilsson, J. Y. Paquet, A. Stipniece, D. A. Stroud, J. Wahl, M. Zenatello & A. Lehtikainen, 2015. Climate-driven changes in winter abundance of a migratory waterbird in relation to EU protected areas. *Diversity and Distributions* 21: 571–582. <https://doi.org/10.1111/DDI.12300>.
- Pinel-Aloul, B. & A. Ghadouani, 2007. Spatial heterogeneity of planktonic microorganisms in aquatic systems. In Franklin, R. B. & A. L. Mills (eds), *The Spatial Distribution of Microbes in the Environment*. Springer, Dordrecht: 203–310.
- Proctor, V. W. & C. R. Malone, 1965. Further evidence of the passive dispersal of small aquatic organisms via the intestinal tract of birds. *Ecology* 46: 728–729.
- Proctor, V. W., C. R. Malone & V. L. DeVlaming, 1967. Dispersal of aquatic organisms: viability of disseminules recovered from the intestinal tract of captive killdeer. *Ecology* 48: 672–676.
- R Core Team, 2023. R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. <https://www.R-project.org/>. Vienna, Austria.
- Ramírez, F., C. Rodríguez, J. Seoane, J. Figuerola & J. Bustamante, 2018. How will climate change affect endangered Mediterranean waterbirds? *PLoS One* 13: e0192702. <https://doi.org/10.1371/JOURNAL.PONE.0192702>.
- Rendón, M. A., A. J. Green, E. Aguilera & P. Almaraz, 2008. Status, distribution and long-term changes in the waterbird community wintering in Doñana, south-west Spain. *Biological Conservation* 141: 1371–1388. <https://doi.org/10.1016/J.BIOCON.2008.03.006>.
- Reynolds, C., N. A. F. Miranda & G. S. Cumming, 2015. The role of waterbirds in the dispersal of aquatic alien and invasive species. *Diversity and Distributions* 21: 744–754. <https://doi.org/10.1111/DDI.12334>.
- Rodrigo, M. A., M. Martín, C. Rojo, S. Gargallo, M. Segura & N. Oliver, 2013a. The role of eutrophication reduction of two small man-made Mediterranean lagoons in the context of a broader remediation system: Effects on water quality and plankton contribution. *Ecological Engineering* 61: 371–382. <https://doi.org/10.1016/J.ECOLENG.2013.09.038>.
- Rodrigo, M. A., C. Rojo, J. L. Alonso-Guillén & P. Vera, 2013b. Restoration of two small Mediterranean lagoons: the dynamics of submerged macrophytes and factors that affect the success of revegetation. *Ecological Engineering* 54: 1–15. <https://doi.org/10.1016/J.ECOLENG.2013.01.022>.
- Rogers, D. C., 2014. Larger hatching fractions in avian dispersed anostracan eggs (Branchiopoda). *Journal of Crustacean Biology* 34: 135–143. <https://doi.org/10.1163/1937240X-00002220>.
- Rueda, J., J. A. Aguilar-Alberola & F. Mezquita-Joanes, 2006. Contribución al conocimiento de los crustáceos (Arthropoda, Crustacea) de las Malladas de la Devesa del Parque Natural de la Albufera (Valencia). *Boletín de la Asociación Española de Entomología* 30: 9–29.
- Sabagh, L. T., R. J. P. Dias, C. W. C. Branco & C. F. D. Rocha, 2011. News records of phoresy and hyperphoresy among treefrogs, ostracods, and ciliates in bromeliad of Atlantic forest. *Biodiversity and Conservation* 20: 1837–1841. <https://doi.org/10.1007/S10531-011-0050-Z>.

- Santamaría, L., I. Charalambidou, D. Viana & E. van Donk, 2023. Evidence that long-distance dispersal of aquatic invertebrates by ducks increases with propagule size. *Freshwater Biology* 68: 1530–1541. <https://doi.org/10.1111/fwb.14146>.
- Schmit, O., G. Rossetti, J. Vandekerckhove & F. Mezquita, 2007. Food selection in *Eucypris virens* (Crustacea: Ostracoda) under experimental conditions. *Hydrobiologia* 585: 135–140. <https://doi.org/10.1007/s10750-007-0634-5>.
- SEO BirdLife, 2021a. Seguimiento de la avifauna invernante en l'Albufera de València (2020–21). Valencia.
- SEO BirdLife, 2021b. Seguimiento de las aves acuáticas nidificantes y macroinvertebrados acuáticos durante el cultivo de arroz en l'Albufera de Valencia (2021). Valencia.
- Silva, G. G., A. J. Green, C. Stenert & L. Maltchik, 2021. Invertebrate dispersal by waterbird species in neotropical wetlands. *Instituto Internacional de Ecologia. Brazilian Journal of Biology* 84: e250280. <https://doi.org/10.1590/1519-6984.250280>.
- Soons, M. B., A. L. Brochet, E. Kleyheeg & A. J. Green, 2016. Seed dispersal by dabbling ducks: an overlooked dispersal pathway for a broad spectrum of plant species. *Journal of Ecology* 104: 443–455. <https://doi.org/10.1111/1365-2745.12531>.
- Soto, I., P. Balzani, L. Carneiro, R. N. Cuthbert, R. Macêdo, A. Serhan Tarkan, D. A. Ahmed, A. Bang, K. Bacela-Spychalska, S. A. Bailey, T. Baudry, L. Ballesteros-Mejia, A. Bortolus, E. Briski, J. R. Britton, M. Buřič, M. Camacho-Cervantes, C. Cano-Barbacil, D. Copilaș-Ciocianu, N. E. Coughlan, P. Courtois, Z. Csabai, T. Dalu, V. De Santis, J. W. E. Dickey, R. D. Dimarco, J. Falk-Andersson, R. D. Fernandez, M. Florencio, A. C. S. Franco, E. García-Berthou, D. Giannetto, M. M. Glavendekic, M. Grabowski, G. Heringer, I. Herrera, W. Huang, K. L. Kamelamela, N. I. Kirichenko, A. Kouba, M. Kourantidou, I. Kurtul, G. Laufer, B. Lipták, C. Liu, E. López-López, V. Lozano, S. Mammola, A. Marchini, V. Meshkova, M. Milardi, D. L. Musolin, M. A. Nuñez, F. J. Oficialdegui, J. Patoka, Z. Pattison, D. Pincheira-Donoso, M. Piria, A. F. Probert, J. J. Rasmussen, D. Renault, F. Ribeiro, G. Rilov, T. B. Robinson, A. E. Sanchez, E. Schwindt, J. South, P. Stoett, H. Verreycken, L. Vilizzi, Y. Wang, Y. Watari, P. M. Wehi, A. Weiperth, P. Wiberg-Larsen, S. Yapıcı, B. Yoğurtcuoğlu, R. D. Zenni, B. S. Galil, J. T. A. Dick, J. C. Russell, A. Ricciardi, D. Simberloff, C. J. A. Bradshaw & P. J. Haubrock, 2024. Taming the terminological tempest in invasion science. *Biological Reviews* 99: 1357–1390. <https://doi.org/10.1111/brv.13071>.
- Spencer, M. & L. Blaustein, 2001. Hatching responses of temporary pool invertebrates to signals of environmental quality. *Israel Journal of Zoology* 47: 397–418. <https://doi.org/10.1092/23F2-2XBW-252B-DU5C>.
- Valls, L., A. Castillo-Escrivà, L. Barrera, E. Gómez, J. A. Gil-Delgado, F. Mesquita-Joanes & X. Armengol, 2017. Differential endozoochory of aquatic invertebrates by two duck species in shallow lakes. *Acta Oecologica* 80: 39–46. <https://doi.org/10.1016/j.actao.2017.03.003>.
- Valls, L., A. Castillo-Escrivà, F. Mesquita-Joanes & X. Armengol, 2016. Human-mediated dispersal of aquatic invertebrates with waterproof footwear. *Ambio* 45: 99–109. <https://doi.org/10.1007/s13280-015-0689-x>.
- Vandekerckhove, J., B. Niessen, S. Declerck, E. Jeppesen, J. M. Conde Porcuna, L. Brendonck & L. De Meester, 2004. Hatching rate and hatching success with and without isolation of zooplankton resting stages. *Hydrobiologia* 526: 235–241. <https://doi.org/10.1023/B:HYDR.0000041598.68424.FC>.
- van Leeuwen, C. H. A., M. B. Soons, L. G. V. T. I. Vandionant, A. J. Green & E. S. Bakker, 2023. Seed dispersal by waterbirds: a mechanistic understanding by simulating avian digestion. *Ecography*. <https://doi.org/10.1111/ECOG.06470>.
- van Leeuwen, C. H. A., G. van der Velde, B. van Lith & M. Klaassen, 2012. Experimental quantification of long distance dispersal potential of aquatic snails in the gut of migratory birds. *PLoS One* 7: e32292. <https://doi.org/10.1371/JOURNAL.PONE.0032292>.
- Vanschoenwinkel, B., A. Waterkeyn, T. Nihwatiwa, T. Pinceel, E. Spooren, A. Geerts, B. Clegg & L. Brendonck, 2011. Passive external transport of freshwater invertebrates by elephant and other mud-wallowing mammals in an African savannah habitat. *Freshwater Biology* 56: 1606–1619. <https://doi.org/10.1111/j.1365-2427.2011.02600.x>.
- Vanschoenwinkel, B., A. Waterkeyn, T. Vandecaetsbeek, O. Pineau, P. Grillas & L. Brendonck, 2008. Dispersal of freshwater invertebrates by large terrestrial mammals: a case study with wild boar (*Sus scrofa*) in Mediterranean wetlands. *Freshwater Biology* 53: 2264–2273. <https://doi.org/10.1111/j.1365-2427.2008.02071.x>.
- Viain, A., F. Corre, P. Delaporte, E. Joyeux & P. Bocher, 2011. Numbers, diet and feeding methods of Common Shelduck *Tadorna tadorna* wintering in the estuarine bays of Aiguillon and Marennes-Ol. *Wildfowl* 61: 121–141.
- Walmsley, J. G. & M. E. Moser, 1981. The winter food and feeding habits of Shelduck in the Camargue, France. *Wildfowl* 32: 99–106.
- Waterkeyn, A., B. Vanschoenwinkel, S. Elsen, M. Anton-Pardo, P. Grillas & L. Brendonck, 2010. Unintentional dispersal of aquatic invertebrates via footwear and motor vehicles in a Mediterranean wetland area. *Aquatic Conservation: Marine and Freshwater Ecosystems*. 20: 580–587. <https://doi.org/10.1002/aqc.1122>.

Publisher's Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.