

Journal section: Endodontics

Publication Types: Research

doi:10.4317/medoral.17680

<http://dx.doi.org/doi:10.4317/medoral.17680>

Antimicrobial activity and *enterococcus faecalis* biofilm formation on chlorhexidine varnishes

María-Teresa Arias-Moliz ¹, Carmen-María Ferrer-Luque ², María-Paloma González-Rodríguez ³, Esther Navarro-Escobar ⁴, Márcia-Furtado-Antunes de Freitas ⁵, Pilar Baca ⁶

¹ DDS, PhD, Assistant Professor, Department of Microbiology, School of Dentistry. University of Granada, Campus de Cartuja, Colegio Máximo s/n, Granada, Spain

² DDS, MD, PhD, Associate Professor Department of Preventive and Operative Dentistry, School of Dentistry. University of Granada, Campus de Cartuja, Colegio Máximo s/n, Granada, Spain

³ DDS, PhD, Associate Professor, Department of Preventive and Operative Dentistry, School of Dentistry. University of Granada, Campus de Cartuja, Colegio Máximo s/n, Granada, Spain

⁴ BDS, Postgraduate Student, Department of Preventive and Operative Dentistry, School of Dentistry. University of Granada, Campus de Cartuja, Colegio Máximo s/n, Granada, Spain

⁵ DDS, Postgraduate Student, Department of Dental Materials, School of Dentistry. University of Sao Paulo, Rua Barão do Triunfo, Sao Paulo, Brazil

⁶ DDS, MD, PhD: Professor, Department of Preventive and Operative Dentistry, School of Dentistry. University of Granada, Campus de Cartuja, Colegio Máximo s/n, Granada, Spain

Correspondence:

Department of Microbiology, School of Dentistry

Campus de Cartuja, Colegio Máximo s/n

E-18071, Granada (Spain)

mtarias@ugr.es

Arias-Moliz MT, Ferrer-Luque CM, González-Rodríguez MP, Navarro-Escobar E, de Freitas MFA, Baca P. Antimicrobial activity and *enterococcus faecalis* biofilm formation on chlorhexidine varnishes. Med Oral Patol Oral Cir Bucal. 2012 Jul 1;17 (4):e705-9.

<http://www.medicinaoral.com/medoralfree01/v17i4/medoralv17i4p705.pdf>

Received: 04/05/2011

Accepted: 12/06/2011

Article Number: 17680 <http://www.medicinaoral.com/>
© Medicina Oral S. L. C.I.F. B 96689336 - pISSN 1698-4447 - eISSN: 1698-6946
eMail: medicina@medicinaoral.com

Indexed in:

Science Citation Index Expanded
Journal Citation Reports
Index Medicus, MEDLINE, PubMed
Scopus, Embase and Emcare
Indice Médico Español

Abstract

Objective: To evaluate, *in vitro*, the antimicrobial activity and biofilm formation of three chlorhexidine varnishes in four *Enterococcus faecalis* strains: *E. faecalis* ATCC 29212, *E. faecalis* EF-D1 (from failed endodontic treatment), *E. faecalis* 072 (cheese) and *E. faecalis* U-1765 (nosocomial infection), and one *Enterococcus durans* strain (failed endodontic treatment).

Study Design: The direct contact test was used to study the antimicrobial activity. Bacterial suspensions were exposed for one hour to EC40, Cervitec (CE) and Cervitec Plus (CEP) varnishes. "Eradication" was defined as 100% bacterial kill. The formation of enterococci biofilms was tested on the surface of the varnishes after 24 hours of incubation and expressed as percentage of biofilm reduction.

Results: EC40 eradicated all strains except *E. faecalis* ATCC 29212, where 98.78% kill was achieved. CE and CEP showed antimicrobial activity against all the strains, but most clearly against *E. durans* and *E. faecalis* 072. EC40 completely inhibited the formation of biofilm of *E. faecalis* ATCC 29212, *E. faecalis* 072 and *E. durans*. CE and CEP led to over 92% of biofilm reduction, except in the case of *E. faecalis* U-1765 on CEP (76.42%).

Conclusion: The three varnishes studied were seen to be effective in killing the tested strains of enterococci and in inhibiting the formation of biofilm, the best results being observed with EC40.

Key words: Biofilm, chlorhexidine varnish, direct contact test, *Enterococcus durans*, *Enterococcus faecalis*, intracanal medication.

Introduction

An important aim of endodontic therapy is the elimination of microorganisms from root canal systems. This may be accomplished using mechanical instrumentation and chemical irrigation, and intracanal medication for persistent infections (1).

Chlorhexidine (CHX) is a cationic molecule with a wide antimicrobial spectrum against both Gram positive and Gram negative bacteria (2). These would include *Enterococcus faecalis* (3), a microorganism frequently isolated from necrotic and failed endodontic treatments (4). Its antibacterial activity and substantivity (5) make CHX widely used as a root canal irrigant (6,7) and intracanal medication (8) and it is known to reduce and/or delay the entry of bacteria into the root canal system (9).

Available in different formulations, CHX is commonly used in endodontics as a solution or gel (2). CHX has recently been incorporated into a variety of sustained-release systems, including varnishes, in order to prolong the period of active-agent delivery and achieve maximum antimicrobial effectiveness (10). Some of these slow-release systems have proven effective as intracanal medication against *E. faecalis* (11-13). The CHX varnishes apparently penetrate and seal tubules in dentin (14), reducing the level of mutans streptococci in exposed root surfaces (15) and it may help control established root caries lesions (16). However, its antimicrobial activity against bacteria involved in root canal infections is not known. The aim of this study was therefore to evaluate, in vitro, the antimicrobial activity and the ability to reduce biofilm formation of three CHX varnishes, two of them containing thymol, against four *E. faecalis* strains and one *E. durans* strain.

Material and Methods

-Microorganisms and tested materials

The bacterial strains used in the study and their source are listed in table 1. Bacterial strains were taken from a 4°C stock culture and streaked out twice on BHI (Scharlau Chemie S.A., Barcelona, Spain) agar plates for 24 hours at 37°C. Colonies were suspended in BHI to obtain a 1 McFarland initial bacterial suspension of approximately 3×10^8 colony-forming units per mL (CFU/mL). All strain cultures were checked for purity by Gram stain and colony morphology.

The CHX varnishes tested were: EC40 (35% CHX, 27% sandarac, 38% ethanol; Biodent BV, Nijmegen, The Netherlands), Cervitec (CE; 1% CHX, 1% thymol, etha-

Table 1. Enterococci species and strains and their source.

Bacteria source	Species	Strains
Collection	<i>E. faecalis</i>	ATCC 29212
Failed endodontic treatment	<i>E. faecalis</i>	EF-D1 ¹
Failed endodontic treatment	<i>E. durans</i>	ED-C1 ¹
Goat's milk cheese	<i>E. faecalis</i>	072 ²
Human nosocomial infection	<i>E. faecalis</i>	U-1765 ²

¹ Microbiology Laboratory, School of Dentistry, University of Granada.

² Microbiology Laboratory, School of Science, University of Granada.

nol, ethyl acetate, polymer; Ivoclar-Vivadent, Schaan, Liechtenstein) and Cervitec Plus (CEP; 1% CHX, 1% thymol, ethanol, water, acrylate and vinylacetate-copolymer; Ivoclar-Vivadent, Schaan, Liechtenstein).

-Direct Contact Test (DCT)

The DCT used to assess the antimicrobial activity of the CHX varnishes is based on previously reported methodology (17,18). While holding a 96-well microtiter plate (Nunc Delta Surface; Nunc, Roskilde, Denmark) vertically, an area of established dimensions on one side of the wells was coated with an equal amount of each varnish using a cavity liner applicator. The varnishes were applied following the manufacturer's instructions, and were allowed to dry for 30 minutes.

A 10- μ L aliquot of the initial bacterial suspension was placed on the surface of each varnish. Bacterial suspensions placed on the wall of uncoated wells served as the positive control. After incubation for 1 hour at 37°C with 95% relative humidity to ensure direct contact between bacteria and tested materials, 240 μ L of sterile BHI was added to each well. The bacterial suspension was mixed for 1 minute, diluted serially and plated for viable cell counting. Ten replicates per strain and varnish were performed.

-Biofilm Formation Test (BFT)

The biofilm model used was the MBEC high-throughput (HTP) device (Innovotech, Edmonton, Alberta, Canada) (3,19,20). This batch-culture apparatus has a lid with 96 pegs that fits over a standard 96-well microtiter plate (21). Ten pegs were coated with each varnish, following the manufacturer's instructions; and 10 uncoated pegs served as the positive and sterility controls. Each assay was performed for a total of ten replicates per strain and CHX varnish in different devices.

The wells of the microtiter plate were inoculated with 150 μ L of a 1 in 30 dilution of the initial bacterial suspension, while 10 wells were inoculated with sterile BHI for the

sterility control. The coated peg lid was fitted inside the wells, and the device was placed on a rocking table (Swing Sw 8 10000-00015. OVAN, Badalona, Spain) at 5 rocks per minute for 24 hours of incubation at 37°C. Biofilms forming on the pegs were rinsed twice with 0.9% saline solution for 2 minutes to remove loosely adherent planktonic bacteria. The lid was then transferred to a microtiter recovery plate with 200 µL of BHI/well and sonicated on a water-table sonicator (Model 5510E–MT; Branson, Danbury, CT) for 10 minutes to disrupt the biofilm structure. The viability of the biofilms was determined by spot plating 10-µL aliquots of recovery biofilms onto BHI agar, then incubating for 24 hours at 37°C.

The antimicrobial activity and the capacity to reduce biofilm formation of each CHX varnish on the strains tested were determined by calculating the percentage of reduction of viable bacteria as follows: $[1 - (\text{mean CFU}_{\text{CHX varnish}} / \text{mean CFU}_{\text{initial bacterial number}})] \times 100$. In the DCT, the term ‘eradication’ was used to denote the death of 100% of the bacterial population. The formation of enterococci biofilms was expressed as percentage of biofilm reduction with respect to the control.

To compare the efficacies of the different CHX varnishes and the strains tested when the percentage kill varied

from 100%, the Student t-test was used, previously subjecting data to the Anscombe transformation.

Results

The results of the antibacterial effects of the CHX varnishes from the DCT are listed in table 2. EC40 showed the best antibacterial action, eradicating all the enterococci strains except for *E. faecalis* ATCC 29212 (98.78%). CE and CEP, while variable in effectivity, produced the greatest kill percentages of strains *E. faecalis* 072 and *E. durans* ED-C1.

No significant differences were obtained when the effects of CE were compared with those of CEP against *E. faecalis* ATCC 29212 (p=0.549) or *E. faecalis* EF-D1 (p=0.705); but differences were indeed significant for the rest of the bacteria tested.

Table 3 shows the percentage of reduction of biofilm formation. *E. faecalis* 072 formed no biofilm on any of the three varnishes tested (100% reduction); *E. faecalis* ATCC 29212 and *E. durans* ED-C1 were likewise unable to create biofilm upon EC40. The percentage of biofilm reduction on CE and CEP was over 92% in all cases except for *E. faecalis* U-1765 on CEP, which gave a mean value of 76.42%.

Table 2. Percentage of kill of five enterococci strains after direct contact with chlorhexidine varnishes (mean±SD).

	EC40	Cervitec	Cervitec Plus
<i>E. faecalis</i> ATCC 29212	98.78±0.01	36.17±0.13 ^{a,b,1}	32.55±0.16 ^{a,b,1}
<i>E. faecalis</i> EF-D1	100.00±0.00 ^a	46.86±0.22 ^{a,1}	42.85±0.11 ^{a,1}
<i>E. faecalis</i> 072	100.00±0.00 ^a	77.59±0.03	83.08±0.05
<i>E. faecalis</i> U-1765	100.00±0.00 ^a	37.22±0.09 ^b	18.51±0.18 ^b
<i>E. durans</i> ED-C1	100.00±0.00 ^a	88.46±0.06	99.95±0.00

Read vertically, same letters indicate non-significant differences.
Read horizontally, same numbers indicate non-significant differences.

Table 3. Percentage of reduction of biofilm formation of four *E. faecalis* strains and one *E. durans* strain by chlorhexidine varnishes (mean±SD).

	EC40	Cervitec	Cervitec Plus
<i>E. faecalis</i> ATCC 29212	100.00±0.00 ^a	95.57±0.05 ^{a,1}	98.71±0.01 ^{a,1}
<i>E. faecalis</i> EF-D1	99.98±0.00 ^{b,1}	98.70±0.01 ^{b,1}	92.55±0.04
<i>E. faecalis</i> 072	100.00±0.00 ^{a,1}	100.00±0.00 ¹	100.00±0.00 ¹
<i>E. faecalis</i> U-1765	99.03±0.01 ^{b,1}	98.39±0.03 ^{b,1}	76.42±0.22
<i>E. durans</i> ED-C1	100.00±0.00 ^a	92.69±0.04 ^a	96.69±0.05 ^a

Read vertically, same letters indicate non-significant differences.
Read horizontally, same numbers indicate non-significant differences.

Discussion

The use of intracanal medication can help eliminate the bacteria remaining after chemo-mechanical instrumentation and may prevent microorganism invasion between treatment appointments (1). Our results suggest that all three varnishes studied exert antimicrobial activity against the five strains tested, and are capable of preventing or reducing the formation of biofilm.

Although the most frequently used intracanal medication is calcium hydroxide, when CHX is used, *E. faecalis* is better eliminated from dentinal tubules (5). As inter-appointment medication, it may be applied in the form of a gel (8) or incorporated into sustained-release devices (13). The potential benefits of CHX varnishes in endodontics may stem from a greater and more prolonged release from dental tissue than with CHX gels (22), and their ability to penetrate and disinfect dentin tubules of the surface root (14).

The bacteria selected for our study were enterococci strains, particularly *E. faecalis*, which are often isolated from necrotic or improperly filled root canal systems (23). *E. faecalis* ATCC 29212 is a strain of reference widely used in antimicrobial susceptibility studies (13). Moreover, two enterococci strains isolated from failed endodontic treatments were studied, *E. faecalis* EF-D1 and *E. durans* ED-C1, the latter reportedly present in root canals (24). The origin of enterococci found in the oral cavity is unclear; and whereas the most likely source would be food (25), nosocomial infections are another possibility. We therefore used strains pertaining to both sources: *E. faecalis* 072 (26) and *E. faecalis* U-1765.

A DCT (17) with several modifications was used to evaluate the antimicrobial activity of the varnishes, since it permits a direct evaluation of their bactericidal effects. This method is quantitative and reliable, and reproduces the contact of the test microorganism with the varnishes. Our results showed that EC40 was the most effective CHX varnish, eradicating all the strains tested, except *E. faecalis* ATCC 29212. CE and CEP gave the highest kill percentages in contact with *E. durans* ED-C1 (over 88%) and *E. faecalis* 072 (over 77%), CEP proving to be significantly more effective than CE. Neither varnish produced kill over 50% against the rest of the strains. This may be due to the lesser concentration of the active ingredients in these preparations, and to the presence of polymers or copolymers in their formulation, which may limit their diffusion.

For the BFT, the MBEC-HTP device was considered appropriate because it allows for the simultaneous formation of 96 statistically equivalent biofilms under similar conditions (19, 21). After 24 hours of incubation (3), the results of the positive controls showed each strain to have a different capacity for forming biofilm. *E. faecalis* 072 created a lesser amount of biofilm, which would explain its 100% reduction with the three varnishes. The two

strains isolated from root canals, *E. durans* ED-C1 followed by *E. faecalis* EF-D1, formed a great amount of biofilm. This finding contrasts with the results reported by Duggan and Sedgley (27), who showed *E. faecalis* strains from endodontic sources to have a lower inherent capacity for biofilm formation.

In parallel to the results seen for DCT, EC40 was the most effective varnish, able to fully inhibit biofilm formation of three strains. This may be attributed to its high concentration of CHX, which would provide greater substantive antimicrobial activity (28). Lee et al (13) similarly obtained strong antimicrobial action on contaminated dentin blocks with *E. faecalis* using a polymeric CHX controlled device with 40% CHX. CE and CEP exhibited a lesser capacity to inhibit biofilm formation in our study, which also varied depending on the species involved. The lower concentration of CHX (1%), even though it included thymol in its composition (29), might explain these results. However, both these varnishes achieved a percentage of reduction over 92%; in contrast, CEP was not very effective against *E. faecalis* U-1765 (76.42%). There is no clear evidence pointing to one of these substances as better than the other, a point brought out previously in the context of cariogenic bacteria (30).

The study described here demonstrates that the CHX varnishes —and particularly EC40— exert effective antimicrobial action against the enterococci strains tested, and that they are able to inhibit or considerably reduce the formation of biofilms of these bacteria. Despite the limitations of this in vitro study, we may state that the results are promising, and encourage further studies to evaluate other properties of varnishes and new sustained-release systems.

References

1. Siqueira JF Jr, Rôças IN. Clinical implications and microbiology of bacterial persistence after treatment procedures. *J Endod.* 2008;34:1291-1301.
2. Mohammadi Z, Abbott PV. The properties and applications of chlorhexidine in endodontics. *Int Endod J.* 2009;42:288-302.
3. Arias-Moliz MT, Ferrer-Luque CM, Espigares-García M, Baca P. *Enterococcus faecalis* biofilms eradication by root canal irrigants. *J Endod.* 2009;35:711-4.
4. Rôças IN, Siqueira JF Jr, Santos KR. Association of *Enterococcus faecalis* with different forms of periradicular diseases. *J Endod.* 2004;30:315-20.
5. Athanassiadis B, Abbott PV, Walsh LJ. The use of calcium hydroxide, antibiotics and biocides as antimicrobial medicaments in endodontics. *Aust Dent J.* 2007;52:S64-82.
6. Komorowski R, Grad H, Wu XY, Friedman S. Antimicrobial substantivity of chlorhexidine-treated bovine root dentin. *J Endod.* 2000;26:315-7.
7. Siqueira JF Jr, Paiva SS, Rôças IN. Reduction in the cultivable bacterial populations in infected root canals by a chlorhexidine-based antimicrobial protocol. *J Endod.* 2007;33:541-7.
8. Malkhassian G, Manzur AJ, Legner M, Fillery ED, Manek S, Basrani BR, et al. Antibacterial efficacy of MTAD final rinse and two percent chlorhexidine gel medication in teeth with apical

- periodontitis: a randomized double-blinded clinical trial. *J Endod.* 2009;35:1483-90.
9. Gomes BP, Sato E, Ferraz CC, Teixeira FB, Zaia AA, Souza-Filho FJ. Evaluation of time required for recontamination of coronally sealed canals medicated with calcium hydroxide and chlorhexidine. *Int Endod J.* 2003;36:604-9.
 10. Puig Silla M, Montiel Company JM, Almerich Silla JM. Use of chlorhexidine varnishes in preventing and treating periodontal disease. A review of the literature. *Med Oral Patol Oral Cir Bucal.* 2008;13:E257-60.
 11. Heling I, Sommer M, Steinberg D, Friedman M, Sela MN. Microbiological evaluation of the efficacy of chlorhexidine in a sustained-release device for dentine sterilization. *Int Endod J.* 1992;25:15-9.
 12. Huang J, Wong HL, Zhou Y, Wu XY, Grad H, Komorowski R, et al. In vitro studies and modeling of a controlled-release device for root canal therapy. *J Control Release.* 2000;67:293-307.
 13. Lee Y, Han SH, Hong SH, Lee JK, Ji H, Kum KY. Antimicrobial efficacy of a polymeric chlorhexidine release device using in vitro model of *Enterococcus faecalis* dentinal tubule infection. *J Endod.* 2008;34:855-8.
 14. Arends J, Duschner H, Ruben JL. Penetration of varnishes into demineralized root dentine in vitro. *Caries Res.* 1997;31:201-5.
 15. Wicht MJ, Haak R, Lummert D, Noack MJ. Treatment of root caries lesions with chlorhexidine-containing varnishes and dentin sealants. *Am J Dent.* 2003;16:25A-30A.
 16. Baca P, Clavero J, Baca AP, González-Rodríguez MP, Bravo M, Valderrama MJ. Effect of chlorhexidine-thymol varnish on root caries in a geriatric population: a randomized double-blind clinical trial. *J Dent.* 2009;37:679-85.
 17. Weiss EI, Shalhav M, Fuss Z. Assessment of antibacterial activity of endodontic sealers by a direct contact test. *Endod Dent Traumatol.* 1996;12:179-84.
 18. Zhang H, Shen Y, Ruse ND, Haapasalo M. Antibacterial activity of endodontic sealers by modified direct contact test against *Enterococcus faecalis*. *J Endod.* 2009;35:1051-5.
 19. Ceri H, Olson ME, Stremick C, Read RR, Morck D, Buret A. The Calgary Biofilm Device: new technology for rapid determination of antibiotic susceptibilities of bacterial biofilms. *J Clin Microbiol.* 1999;37:1771-6.
 20. Arias-Moliz MT, Ferrer-Luque CM, González-Rodríguez MP, Valderrama MJ, Baca P. Eradication of *Enterococcus faecalis* biofilms by cetrime and chlorhexidine. *J Endod.* 2010;36:87-90.
 21. Harrison JJ, Turner RJ, Ceri H. High-throughput metal susceptibility testing of microbial biofilms. *BMC Microbiol.* 2005;3:53.
 22. Attin T, Abouassi T, Becker K, Wiegand A, Roos M, Attin R. A new method for chlorhexidine (CHX) determination: CHX release after application of differently concentrated CHX-containing preparations on artificial fissures. *Clin Oral Investig.* 2008;12:189-96.
 23. Subramanian K, Mickel AK. Molecular analysis of persistent periradicular lesions and root ends reveals a diverse microbial profile. *J Endod.* 2009;35:950-7.
 24. Mejäre B. *Streptococcus faecalis* and *Streptococcus faecium* in infected dental root canals at filling and their susceptibility to azidocillin and some comparable antibiotics. *Odontol Revy.* 1975;26:193-204.
 25. Zehnder M, Guggenheim B. The mysterious appearance of enterococci in filled root canals. *Int Endod J.* 2009;42:277-87.
 26. Martín-Platero AM, Valdivia E, Maqueda M, Martínez-Bueno M. Characterization and safety evaluation of enterococci isolated from Spanish goats' milk cheeses. *Int J Food Microbiol.* 2009;132:24-32.
 27. Duggan JM, Sedgley CM. Biofilm formation of oral and endodontic *Enterococcus faecalis*. *J Endod.* 2007;33:815-8.
 28. Davies A. The mode of action of chlorhexidine. *J Periodontal Res Suppl.* 1973;12:68-75.
 29. Huizinga ED, Ruben JL, Arends J. Chlorhexidine and thymol release from a varnish system. *J Biol Buccale.* 1991;19:343-8.
 30. Sköld-Larsson K, Sollenius O, Petersson LG, Twetman S. Effect of topical applications of a novel chlorhexidine-thymol varnish formula on mutans streptococci and caries development in occlusal fissures of permanent molars. *J Clin Dent.* 2009;20:223-6.